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High-Resolution LC-MS Characterization of Antibody Drug Conjugates with PNGase F Enzymatic Cleavage of N-linked Oligosaccharides using Biozen[™] Native-RP-5

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Antibody drug conjugates (ADCs) remain a highly researched biotherapeutic modality due to their success in targeting and killing tumor cells without compromising healthy ones. Cysteine-linked ADCs have improved pharmacokinetics, efficacy and reduced toxicity. Examples of approved cysteine-linked ADCs are Adcetris[®] and Tivdak[®].

Characterization of ADCs is still an ongoing challenge. Hydrophobic Interaction Chromatography (HIC) is a traditional approach but requires sample treatment for MS friendly analysis and although reversed analysis is an acceptable approach, LC conditions cause the antibody subunits with reduced disulfide bonds to dissociate.

In this technical note, we demonstrate the performance of Biozen[™] Native RP-5, a non-porous, hydrophilic particle designed to preserve the native forms of intact biomolecules under reversed phase conditions, for the intact mass measurement resulting from deglycosylation via PNGase F, as well as the separation of intact ADC drug-to-antibody ratio (DAR) species, of four cysteine-linked ADCs.

Sample Preparation

Four commercial cysteine linked ADC drugs were tested: 1) Adcetris (brentuximab vedotin), 2) Padcev[®] (enfortumab vedotin), 3) Polivy[®] (polartuzumab vedotin-piiq), 4) Tivdak (tisotumab vedotin-tftv).

N-linked oligosaccharides of ADC samples were enzymatically cleaved using PNGase F from Promega® (cat# V4831) according to Promega protocol without the use of surfactants.

Enzymatically treated ADC samples were buffer exchanged into 20 mM Ammonium Acetate pH 5.2 using Vivaspin® 500 30 kDa centrifugal concentrators prior to analysis. Buffer exchange was performed to reduce fouling of the MS source by components of the ADC formulation and to improve sensitivity for the DAR0 species which elutes close to the void time of the column. The relatively high concentration of salts and other formulation components will cause MS signal suppression for any components eluting near the void time.

LC Condition	ons		
Column:	Biozen Native-RP-5		
Dimensions:	50 x 2.1 mm		
Part No.:	00B-4800-AN		
Mobile Phase:	A: 50 mM Ammor	nium Acetate in Water	
	B: 25mM Ammon	ium Acetate in 50/50 Water/Isopropanol (v/v)	
Gradient:	Time (min)	% B	
	0	0	
	2	0	
	5	15	
	20	50	
	24	50	
	26	100	
	27	100	
	27.1	0	
	30	0	
Flow Rate:	0.2 mL/min		
Injection Volume:	20 μg ADC on-column		
Temperature:	30 °C		
LC System:	Agilent 1290 Infinity II UHPLC		
Detection:	HRMS, UV (280 nm)		
Detector:	SCIEX® X500B HRMS, Agilent® DAD (UV)		
	following compo JetWeaver V35, thermostat and	It 1290 Infinity II UHPLC system contained the onents: G7120A high speed binary pump with G7167B multisampler, G7116B multicolumn G7117B diode array detector (10mm o plactic mobile phase research ware used	

thermostat and G7117B diode array detector (10mm pathlength). Also, plastic mobile phase reservoirs were used to reduce any metallic impurities (especially sodium and potassium) that may leach from standard borosilicate glass reservoirs. Finally, to ensure ADCs remained in their native, intact state the mobile phase organic composition was limited to a maximum of 25 v/v% IPA except for a brief column cleanup step (50 v/v% IPA) at the end of the gradient.

HRMS Conditions

Ionization Mode: ESI Polarity: Positive Source Temperature: 450 °C Declustering Potential: 200 V (50 V spread) CE ± CE Spread: 5 ± 0V Ion Spray Voltage: 5000 V Scan Range: 1000-8000 m/z





Results and Discussion

The PNGase treated ADC samples compared to the non-treated samples show loss of mass approximate to N-linked glycans commonly associated with monoclonal antibodies (see **Figures 1-4** and **Table 1** below).

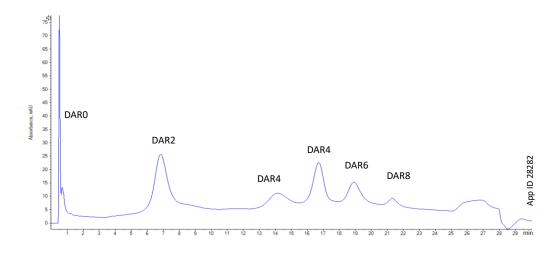
It was also observed that the PNGase treated ADC DAR species showed loss of multiple peaks separated by 162 Da and a simplified mass fingerprint compared to the non-treated samples. This mass shift of 162 Da corresponds to ADC species that have lost N-linked glycans with various glycoforms with added galactose or mannose molecules further confirming that N-linked glycans were cleaved from the ADCs.

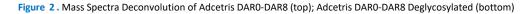
 Table 1. PNGase Treated ADC Samples Comparing Loss of Mass to Approximate

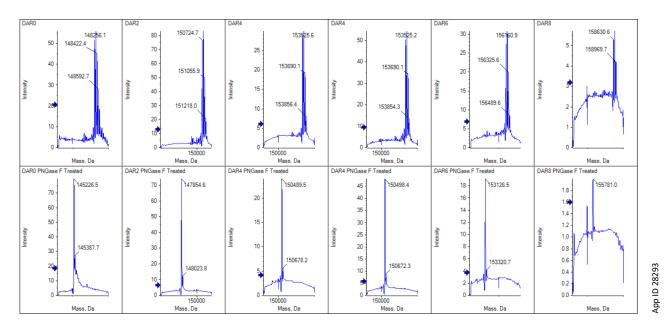
 N-linked Glycans

Sample	Observed Average loss Da	Probable Glycan	Theoretical Average Mass Da
Adcetris®	3030.3	G0F/G1F	3052.8
Padcev®	2875.1	G0F/G0F	2890.7
Polivy®	2858.6	G0F/G0F	2890.7
Tivdak®	2875.0	G0F/G0F	2890.7

Figure 1. A280nm Chromatogram of Adcetris ADC Separation by Biozen™ Native RP-5 Column





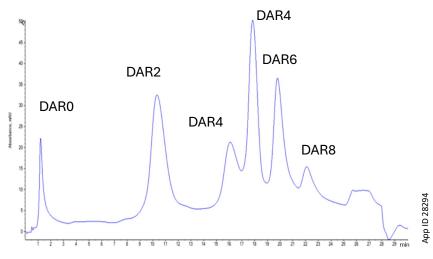


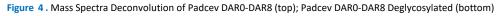
Have questions or want more details on implementing this method? We would love to help! Visit www.phenomenex.com/Chat to get in touch with one of our Technical Specialists



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Figure 3. A280nm Chromatogram of Padcev® ADC Separation by Biozen™ Native RP-5 Column





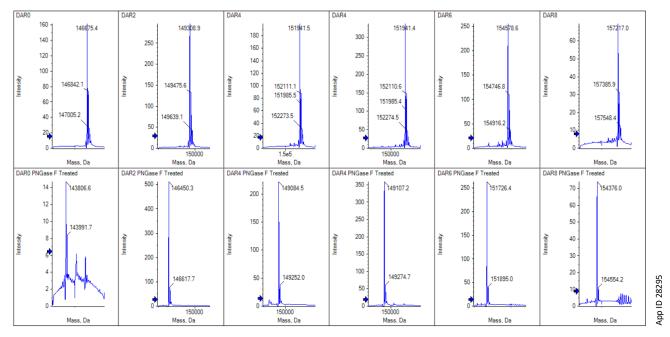
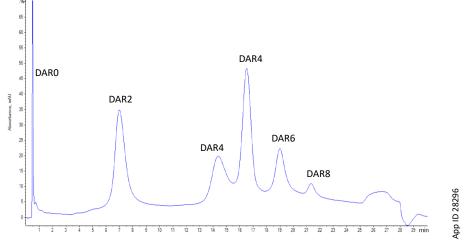


Figure 5 . A280nm Chromatogram of Polivy® ADC Separation by Biozen Native RP-5 Column



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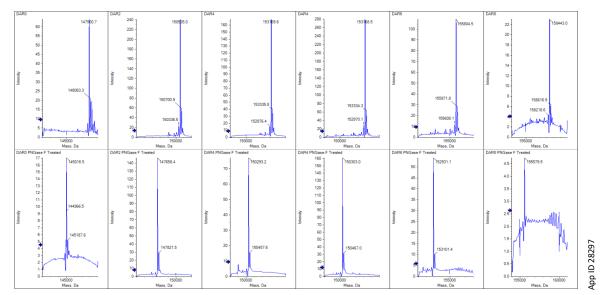
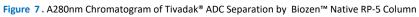


Figure 6. Mass Spectra Deconvolution of Polivy® DAR0-DAR8 (top); Polivy DAR0-DAR8 Deglycosylated (bottom)



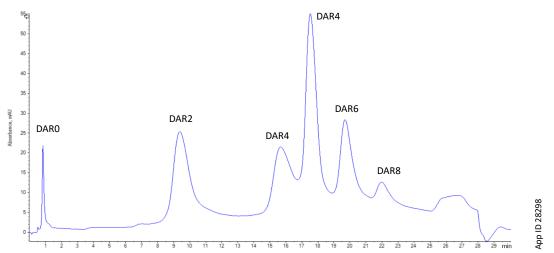
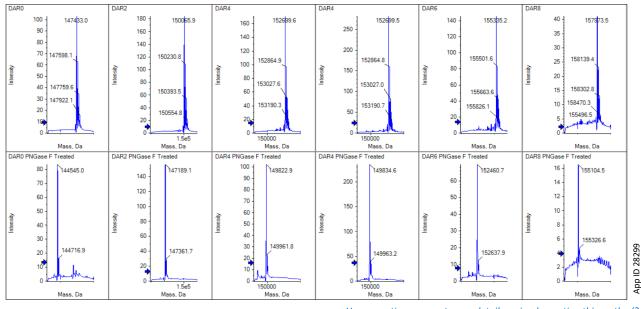


Figure 8. Mass Spectra Deconvolution of Tivadak DAR0-DAR8 (top); Tivadak DAR0-DAR8 Deglycosylated (bottom)



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Conclusion

The challenges in characterizing ADCs necessitate advanced techniques, such as the use of Biozen™ Native RP-5 columns, which enable intact mass measurement under native reverse phase conditions and separation of cysteine linked ADC species without dissociating the antibody subunits. The experimental results confirm successful deglycosylation of N-linked glycans from the ADCs, providing a clearer mass fingerprint and simplifying the analysis. The Biozen Native RP-5 LC column is suitable for LC HRMS analysis of intact ADC for drug-to-antibody ratio (DAR), and PNGase F treated ADCs.

Biozen Ordering Information

Biozen Columns (mm)				
	50 x 2.1	50 x 4.6		
Biozen Native-RP-5	00B-4800-AN	<u>00B-4800-E0</u>		
Biozen Native-RP-1	00B-4799-AN	<u>00B-4799-E0</u>		



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