

TN-1357

Comparing Kinetex™ Biphenyl and Luna™ Omega Polar C18 Columns for Analysis of an 11 Analyte Anticonvulsant Panel

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Introduction

Liquid chromatography with tandem mass spectrometry (LC-MS/MS) methods are considered the gold standard for therapeutic drug monitoring (TDM) of medications. LC-MS/MS provides several advantages for TDM, including high sensitivity, selectivity, and the ability to analyze multiple drugs in a single sample. Typically, serum or whole blood is used as the standard matrix for therapeutic drug monitoring, with serum preferred due to its ease of handling compared to whole blood. However, serum is a complex biological matrix containing various endogenous compounds that can interfere with analysis. These matrix effects can affect the accuracy and precision of drug concentration measurements, often requiring the use of complex sample preparation methods.

In this technical note, we demonstrate a fast and accurate method for the LC-MS/MS analysis of 11 Anticonvulsants using a Strata™-X Method Development 96-well plate that is packed with 4 different polymeric Solid Phase Extraction (SPE) sorbents. This is combined with a fast LC method using a Kinetex 2.6 µm Biphenyl LC column and a Luna Omega 3 µm Polar C18 LC column to resolve all target analytes.

LC Conditions

| Column: | Kinetex 2.6 μm Biphenyl | Luna Omega 3.0 μm Polar C18 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
|--------------------------|--|--|-----|---|----|-----|----|---|----|-----|----|-----|----|------|----|---|----|--|------------|-----|---|----|-----|----|-----|----|-----|----|---|----|-----|----|------|----|---|----|
| Dimensions: | 50 x 3.0 mm | 50 x 3.0 mm | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Part No.: | 00B-4622-Y0 | 00B-4760-Y0 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Mobile Phase: | A: 0.1 % Formic acid in Water B: 0.1 % Formic acid in Methanol | A: 2 mM Ammonium Acetate B: 2 mM Ammonium Acetate in Methanol | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Gradient: | <table><tr><th>Time (min)</th><th>% B</th></tr><tr><td>0</td><td>20</td></tr><tr><td>1.5</td><td>80</td></tr><tr><td>3</td><td>80</td></tr><tr><td>3.1</td><td>95</td></tr><tr><td>3.5</td><td>95</td></tr><tr><td>3.51</td><td>20</td></tr><tr><td>5</td><td>20</td></tr></table> | Time (min) | % B | 0 | 20 | 1.5 | 80 | 3 | 80 | 3.1 | 95 | 3.5 | 95 | 3.51 | 20 | 5 | 20 | <table><tr><th>Time (min)</th><th>% B</th></tr><tr><td>0</td><td>20</td></tr><tr><td>0.5</td><td>20</td></tr><tr><td>1.5</td><td>40</td></tr><tr><td>2.5</td><td>80</td></tr><tr><td>3</td><td>95</td></tr><tr><td>3.5</td><td>95</td></tr><tr><td>3.51</td><td>20</td></tr><tr><td>5</td><td>20</td></tr></table> | Time (min) | % B | 0 | 20 | 0.5 | 20 | 1.5 | 40 | 2.5 | 80 | 3 | 95 | 3.5 | 95 | 3.51 | 20 | 5 | 20 |
| Time (min) | % B | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 0 | 20 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 1.5 | 80 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 3 | 80 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 3.1 | 95 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 3.5 | 95 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 3.51 | 20 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 5 | 20 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Time (min) | % B | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 0 | 20 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 0.5 | 20 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 1.5 | 40 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 2.5 | 80 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 3 | 95 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 3.5 | 95 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 3.51 | 20 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 5 | 20 | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Flow Rate: | 0.5 mL/min | 0.8 mL/min | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Injection Volume: | 5 μL | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Temperature: | 40° C | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| LC System: | Agilent® 1260 Infinity | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Detection: | MS/MS | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Detector: | SCIEX® 6500 Triple Quad™ | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |

MS/MS Conditions

| | |
|----------------------------|-------------------|
| Ion Source: | ESI |
| Polarity: | Positive/Negative |
| Source Temperature: | 450° C |
| GS1: | 55 psi |
| GS2: | 60 psi |
| CUR: | 35 psi |
| IS: | +2500 V or -2500V |

Sample Preparation

A 1 ng/mL standard mix was used for initial LC-MS/MS analysis. A more detailed explanation of the use of the Strata-X Method Development 96-well plate (Part No.: [KSO-8209](#)) to determine the best sample extraction protocol for the Anticonvulsants drug class can be found in [TN-0163](#). Briefly, 500 µL of human serum was spiked with an Anticonvulsants standard mix (1 ng/mL) and extracted using the Strata-X Method Development plate under Neutral, Basic, or Acid loading buffer, with subsequent Neutral, Basic, or Acidic elution. The best results were obtained using the Strata-X-CW sorbent chemistry and the Acid load/Basic elution extraction conditions. After dry-down, samples were reconstituted in 500 µL of initial mobile phase and spiked with 5 ng/mL internal standards. 5 µL of sample was injected onto columns for analysis.

Table 1. MS Transitions.

| Analyte | Q1 Mass (Da) | Q3 Mass (Da) |
|-----------------------|--------------|--------------|
| Pregabalin | 160.1 | 55 |
| Gabapentin | 172 | 137 |
| Levetiracetam | 171 | 126.1 |
| Lamotrigine | 256.1 | 211.1 |
| Felbamate | 178.1 | 117.1 |
| Lacosamide | 251.1 | 108 |
| Zonisamide | 212.9 | 132 |
| Topiramate | 338 | 78 |
| Oxcarbazepine | 253 | 236 |
| Carbamazepine Epoxide | 253.1 | 180.1 |
| Carbamazepine | 237.1 | 194.1 |

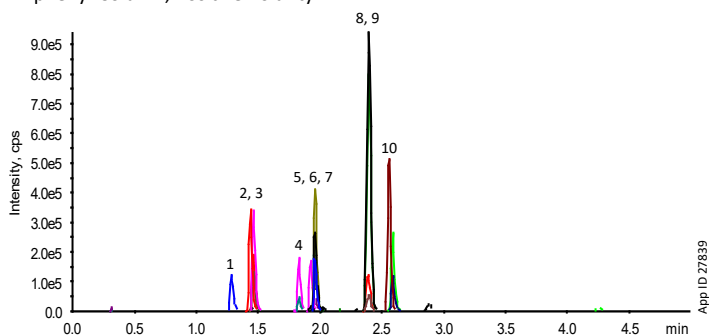


Results and Discussion

A mix of Anticonvulsants standards was analyzed with the Kinetex™ 2.6 µm Biphenyl column showing good peak shape and separation in both positive (Figure 1a) and negative mode (Figure 1b). However, the isobaric analytes Oxcarbazepine and Carbamazepine Epoxide failed to show complete separation (Figure 1c). Because of the lack of separation of this isobaric pair, spiked serum was not analyzed on the Kinetex 2.6 µm Biphenyl column.

The Antipsychotics standards mix was also analyzed with the Luna™ Omega 3 µm Polar C18 column. Analytes were separated with good peak shape (Figures 2a and 2b) and demonstrated great selectivity towards the separation of the critical isobaric pair of Oxcarbazepine and Carbamazepine Epoxide (Figure 2c). To evaluate the chromatography from a matrix-matched sample, 11 Anticonvulsants were spiked into serum and extracted using the Strata™-X Method Development Plate before being analyzed on a Luna Omega 3 µm Polar C18 column. The Strata-X-C sorbent with the acidic load and basic elution solvents were found to be the most successful extraction method for these analytes. For more information on the SPE protocol, please see [TN-0163](#). The same degree of separation of analytes was observed between the extracted serum sample and the neat standard (Figures 3a, 3b, and 3c).

Figure 1a. Analysis of Anticonvulsants Standards on a Kinetex 2.6 µm Biphenyl Column, Positive Polarity.



| Peak No. | Analyte | Retention Time (min) |
|----------|-----------------------|----------------------|
| 1 | Pregabalin | 1.3 |
| 2 | Gabapentin | 1.4 |
| 3 | Levetiracetam | 1.4 |
| 4 | Lamotrigine | 1.8 |
| 5 | Felbamate | 1.9 |
| 6 | Lacosamide | 1.9 |
| 7 | Zonisamide | 1.9 |
| 8 | Oxcarbazepine | 2.4 |
| 9 | Carbamazepine Epoxide | 2.4 |
| 10 | Carbamazepine | 2.6 |

Figure 1b. Analysis of Anticonvulsant Standard Topiramate on a Kinetex 2.6 µm Biphenyl Column, Negative Polarity.

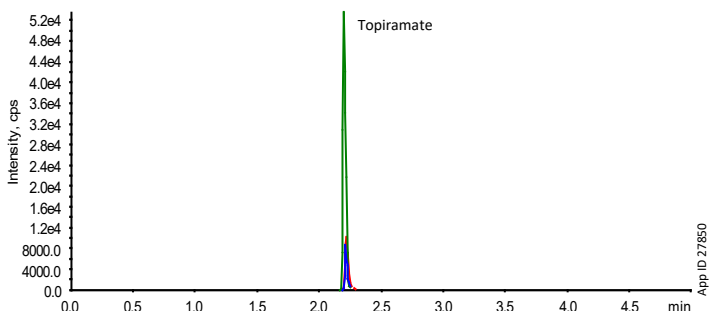


Figure 1c. Incomplete Separation of Isobaric Oxcarbazepine and Carbamazepine Epoxide Anticonvulsants on a Kinetex 2.6 µm Biphenyl Column.

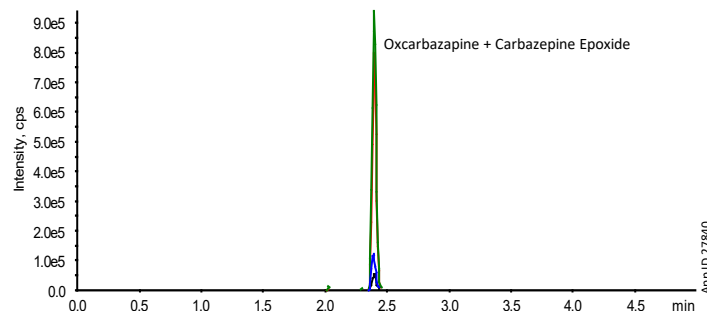
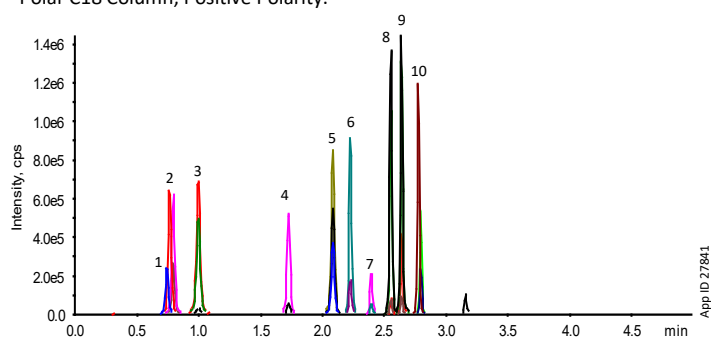


Figure 2a. Analysis of Anticonvulsants Standards on a Luna™ Omega 3 µm Polar C18 Column, Positive Polarity.



| Peak No. | Analyte | Retention Time (min) |
|----------|-----------------------|----------------------|
| 1 | Pregabalin | 0.7 |
| 2 | Gabapentin | 0.8 |
| 3 | Levetiracetam | 1.0 |
| 4 | Zonisamide | 1.7 |
| 5 | Lacosamide | 2.1 |
| 6 | Felbamate | 2.2 |
| 7 | Lamotrigine | 2.4 |
| 8 | Carbamazepine Epoxide | 2.5 |
| 9 | Oxcarbazepine | 2.6 |
| 10 | Carbamazepine | 2.8 |

Figure 2b. Analysis of Anticonvulsant Standard Topiramate on a Luna Omega 3 µm Polar C18 Column, Negative Polarity.

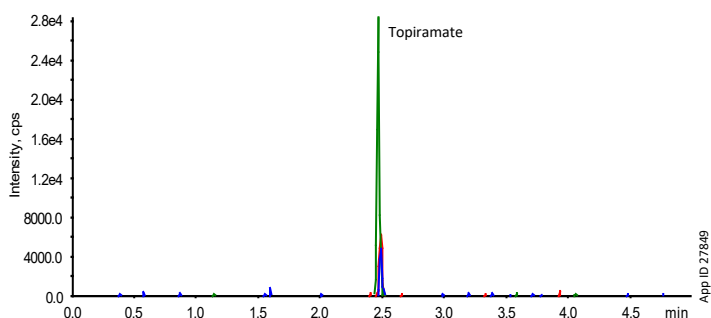


Figure 2c. Separation of Isobaric Oxcarbazepine and Carbamazepine Epoxide Anticonvulsants on a Luna Omega 3 µm Polar C18 Column.

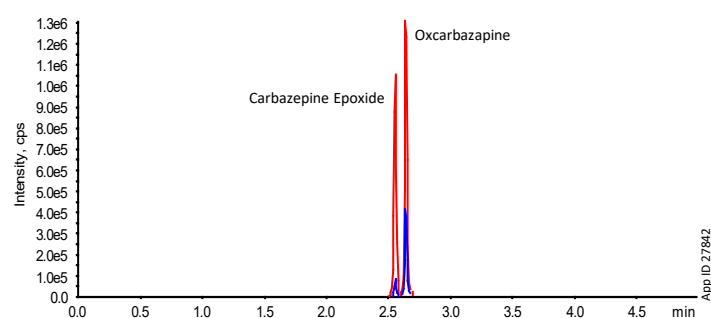
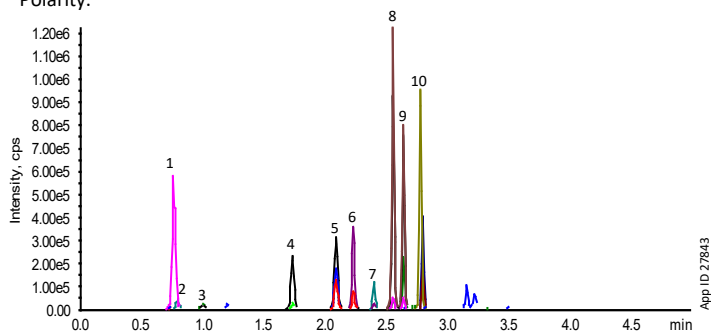


Figure 3a. Analysis of Anticonvulsants Extracted from Serum Using Strata™-X-C, Under Acidic Load and Basic Elution, on a Luna Omega 3 µm Polar C18 Column, Positive Polarity.



| Peak No. | Analyte | Retention Time (min) |
|----------|-----------------------|----------------------|
| 1 | Pregabalin | 0.7 |
| 2 | Gabapentin | 0.8 |
| 3 | Levetiracetam | 1.0 |
| 4 | Zonisamide | 1.7 |
| 5 | Lacosamide | 2.1 |
| 6 | Felbamate | 2.2 |
| 7 | Lamotrigine | 2.4 |
| 8 | Carbamazepine Epoxide | 2.5 |
| 9 | Oxcarbazepine | 2.6 |
| 10 | Carbamazepine | 2.8 |

Figure 3b. Analysis of Anticonvulsant Topiramate Extracted from Serum Using Strata-X-C, Under Acidic Load and Basic Elution, on a Luna Omega 3 µm Polar C18 Column, Negative Polarity.

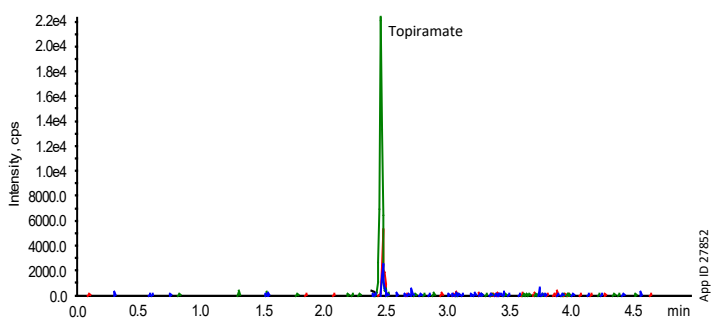
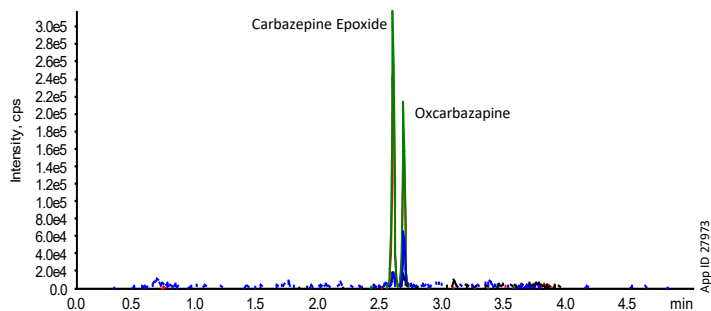


Figure 3c. Separation of Isobaric Oxcarbazepine and Carbamazepine Epoxide Anticonvulsants Extracted from Serum Using Strata-X-C, Under Acidic Load and Basic Elution, on a Luna Omega 3 µm Polar C18 Column.



Conclusions

The developed LC-MS/MS method utilizing the Luna™ Omega 3 µm Polar C18 column has been found to be the optimal chromatography conditions for analysis of the TDM panel of Anticonvulsants. The Method Development plate failed to identify a single optimal protocol for extraction of the Anticonvulsants from serum. Further work is needed to develop an SPE method. A Kinetex™ 2.6 µm Biphenyl column was compared to the Luna Omega 3 µm Polar C18 column but was unable to separate the isobaric pair of Oxcarbazepine and Carbamazepine Epoxide. This suggests that the Luna Omega column is a better option for separating Anticonvulsant analytes when compared to the Kinetex Biphenyl column.

SPE Ordering Information

Strata™-X Method Development 96-Well Plate

| Part No. | Description | Unit |
|--------------------------|--|------|
| KS0-8209 | Strata-X, -X-C, -X-CW, and -X-AW 30 mg/well each | ea |

Kinetex Ordering Information

| 2.6 µm MidBore™ Columns (mm) | | | | SecurityGuard™ ULTRA Cartridges (mm)‡ | | |
|------------------------------|-----------------------------|-----------------------------|-----------------------------|---------------------------------------|-----------------------------|--------------------------|
| Phases | 30 x 3.0 | 50 x 3.0 | 75 x 3.0 | 100 x 3.0 | 150 x 3.0 | 3/pk |
| EVO C18 | 00A-4725-Y0 | 00B-4725-Y0 | — | 00D-4725-Y0 | 00F-4725-Y0 | AJ0-9297 |
| PS C18 | 00A-4780-Y0 | 00B-4780-Y0 | — | 00D-4780-Y0 | 00F-4780-Y0 | AJ0-8950 |
| Polar C18 | — | 00B-4759-Y0 | — | 00D-4759-Y0 | 00F-4759-Y0 | AJ0-9531 |
| Biphenyl | — | 00B-4622-Y0 | — | 00D-4622-Y0 | 00F-4622-Y0 | AJ0-9208 |
| XB-C18 | 00A-4496-Y0 | 00B-4496-Y0 | 00C-4496-Y0 | 00D-4496-Y0 | 00F-4496-Y0 | AJ0-8775 |
| C18 | 00A-4462-Y0 | 00B-4462-Y0 | 00C-4462-Y0 | 00D-4462-Y0 | 00F-4462-Y0 | AJ0-8775 |
| C8 | 00A-4497-Y0 | 00B-4497-Y0 | 00C-4497-Y0 | 00D-4497-Y0 | 00F-4497-Y0 | AJ0-8777 |
| HILIC | 00A-4461-Y0 | — | — | 00D-4461-Y0 | 00F-4461-Y0 | AJ0-8779 |
| Phenyl-Hexyl | — | 00B-4495-Y0 | — | 00D-4495-Y0 | 00F-4495-Y0 | AJ0-8781 |
| F5 | — | 00B-4723-Y0 | — | 00D-4723-Y0 | 00F-4723-Y0 | AJ0-9321 |

for 3.0 mm ID

‡SecurityGuard ULTRA Cartridges require holder, Part No.: [AJ0-9000](#)

Luna Omega Ordering Information

| 3 µm MidBore Columns (mm) | | | SecurityGuard Cartridges (mm) | |
|---------------------------|-----------------------------|-----------------------------|-------------------------------|--------------------------|
| Phases | 50 x 3.0 | 100 x 3.0 | 150 x 3.0 | 4 x 2.0*/10pk |
| Polar C18 | 00B-4760-Y0 | 00D-4760-Y0 | 00F-4760-Y0 | AJ0-7600 |
| PS C18 | 00B-4758-Y0 | 00D-4758-Y0 | 00F-4758-Y0 | AJ0-7605 |
| C18 | 00B-4784-Y0 | 00D-4784-Y0 | 00F-4784-Y0 | AJ0-7611 |
| SUGAR | — | — | 00F-4775-Y0 | AJ0-4496 |

for ID: 2.0 – 3.0 mm

*SecurityGuard Analytical Cartridges require holder, Part No.: [KJ0-4282](#)



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CAUTION: this patent only applies to the analytical-sized guard cartridge holder, and does not apply to SemiPrep, PREP, or ULTRA holders, or to any cartridges.

Strata-X is patented by Phenomenex. U.S. Patent No. 7,119,145.

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